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Endometriosis as a model for inflammation-hormone interactions in ovarian and breast cancers

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ABSTRACT

Chronic inflammation has been implicated in a variety of cancers. In this review, we consider associations between endometriosis and cancers both local (ovarian) and distant (breast). We review the epidemiological data linking endometriosis to ovarian and breast cancers. We then consider evidence for a role for sex steroid hormones and for inflammation in the aetiology of each of these cancers. Finally, we consider that endometriosis may promote alterations in sex steroid hormones and inflammatory mediators. A possible explanation for the association between endometriosis and these reproductive cancers may then be local and systemic enhancement of aberrant inflammatory and hormonal mediators. If this hypothesis is true, endometriosis may need to be considered as a risk factor for ovarian and breast cancers, triggering increasingly intensive surveillance. Moreover, treatments for endometriosis may require consideration of the impact on long-term cancer risk.

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1. Introduction

Chronic inflammation induced by infections and irritants has been implicated in one-sixth of incident cancers world-wide. $^{1-3}$ Just a few examples of non-infectious, chronic inflammants and linked cancers include asbestos fibres and mesothelioma; 4 inflammatory bowel disease and colon cancer; and mastitis and breast cancer. 5 Previously, we proposed that inflammatory exposures may also promote the development of ovarian cancer. 6,7 These examples all involve inflammation and tumour development within the same tissue. Systemic inflammatory diseases and their treatments (e.g., rheumatoid arthritis and TNF- α therapy) have generally been linked to lymphoproliferative, rather than organ-specific malignancies. 8

In this review, we consider associations between endometriosis, defined by the presence of endometrial foci outside the endometrium, and cancers both local (ovarian) and distant (breast). We suggest that endometriosis may promote both tissue-specific and systemic alterations in sex steroid hormones and inflammatory mediators. Thereby, endometriosis may represent a model, wherein inflammatory-hormonal interactions promote both local and distant carcinogenesis.

2. Epidemiological data linking endometriosis, ovarian cancer and breast cancer

Epidemiological studies have consistently supported a link between endometriosis and cancer. Brinton and colleagues

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assessed cancer outcomes among 20,686 women in Sweden who had been hospitalised with endometriosis.9 Cancer outcomes were identified through the National Swedish Cancer Registry after a mean of 11.4 years of follow-up. Significant excess risks were observed for ovarian cancer (standardised incidence ratio 1.9, 95% confidence interval (CI) 1.3-2.8), breast cancer (1.3, 1.1-1.4) and non-Hodgkin's lymphoma (1.8, 1.2-2.6). The risk of ovarian cancer was elevated 4.2-fold among women with a long-standing history of ovarian endometriosis. In a more recent retrospective cohort study conducted in the United States of America (USA), Brinton and colleagues confirmed the relationship between endometriosis and ovarian cancer (standardised incidence ratio 2.5, CI 1.3-4.2), and showed a particularly strong relationship between endometriosis resulting in infertility and ovarian cancer (standardised incidence ratio 4.2, CI 2.0-7.7).¹⁰

Supporting these findings, the 4000 members of the Endometriosis Association completing a health survey¹¹ reported cancer occurrences that were 5-fold higher for ovarian cancer, 6-fold higher for breast cancer, and 2.5-fold higher for non-Hodgkin's lymphoma than expected rates among women in the general population.

Several case-control studies have confirmed the association between endometriosis and ovarian cancer. Ness and colleagues, in a case-control study of 767 women with epithelial ovarian cancer and 1367 controls, after adjustment for age, number of pregnancies, family history of ovarian cancer, race, oral contraceptive use, tubal ligation, hysterectomy and breastfeeding, demonstrated that women with ovarian cancer were 1.7-fold more likely (95% CI 1.2-2.4) to report a history of endometriosis. In a further pooled study of 13,000 women examining the relationships among infertility and ovarian cancer, ovarian cancer was more likely among subfertile women, and particularly among women whose infertility had been diagnosed as resulting from endometriosis (odds ratio (OR) 1.9, 95% CI 1.2-2.9). 12 The only other type of infertility significantly linked to ovarian cancer was infertility of unknown cause (OR 1.5, 95% CI 1.3-1.7), a disorder that hormonal studies suggest may be closely related to endometriosis. 13 The associations between ovarian cancer and either infertility due to endometriosis or infertility due to unknown cause persisted after adjustment for multiple confounding factors. Oral contraceptives, parity and tubal ligation or hysterectomy protected women with endometriosis from developing ovarian cancer. 14

A record-linkage study involving more than 15,000 women in Sweden addressed the link between endometriosis and breast cancer. Although pre-menopausal oophorectomy was associated with a 50% reduction in breast cancer risk, there was a non-significant increase in breast cancer risk among women whose indication for oophorectomy was endometriosis. Moreover, hysterectomy without oophorectomy was associated with an increase in breast cancer risk when the indication was severe endometriosis. Not all epidemiological studies have been positive. Three small retrospective studies designed to assess infertility found no relationship between infertility from endometriosis and ovarian cancer. 16-18 Two of these, however, found relationships between infertility of unknown cause and ovarian cancer. Finally, among women from the Iowa Women's Health Study, self-

reported endometriosis was not associated with ovarian or breast cancer, but was significantly associated with risk for non-Hodgkin's lymphoma. ¹⁹ Nonetheless, most epidemiological studies support a link between endometriosis and cancer, specifically to ovarian and breast cancers.

3. Clinical data linking endometriosis and ovarian cancer

Although, to our knowledge, the only clinical data that demonstrate an association between endometriosis and breast cancer are studies showing that women using tamoxifen for breast cancer may develop endometriosis (presumably on the basis of a pro-oestrogenic effect (see below))^{20,21} there are compelling clinical data relating endometriosis and ovarian cancer. Pathological case series have provided evidence for the transformation of endometriosis into malignancy.^{22–32} Heaps and colleagues noted that 165 published cases showed malignant transformation of endometrial implants, 22 almost 80% occurring within ovarian endometriosis. The overall frequency of malignant transformation was estimated to be 0.3-0.8%.^{22,31} More recent, larger pathology series (up to 1000 cases) have found ovarian cancer in 5-10% of ovarian endometriotic lesions. 29,33 Endometrioid (up to 60% of lesions) and clear-cell (up to 15%) tumours predominate in proportions higher than have been found among ovarian cancers in general (10-20% and 3-10%, respectively),34 demonstrating the distinctive, albeit not exclusive, relationship between endometrioid and clear-cell ovarian cancers and endometriosis. One particularly compelling case report documented a biopsy showing atypia within ovarian endometriosis, followed 3 years later by clear-cell ovarian carcinoma in a rebiopsy from the same ovary.²⁶

With regard to endometriosis occurring within ovarian cancers, Sainz de la Cuesta and colleagues found endometriosis among about 40% of women with stage I endometrioid or clear-cell ovarian carcinoma, about one-third of which represented carcinomas arising out of endometriosis. ²³ Similar results have been reported by others. Endometriosis has generally been found in 21–54% of clear-cell and endometrioid tumours and in 3–9% of serous, mucinous and other tumours. ^{24,27,35,36} Overall, then, pathology and clinical data strongly support the idea that malignant transformation may occur in perhaps 5–10% of women found at surgery to have ovarian endometriosis. In comparison, the general population lifetime risk of ovarian cancer is only 1.5%.

4. Inflammation and cancer

How might endometriosis elevate the risk of both local and distant cancers? We propose a mechanism influenced by inflammation and steroid hormones. Chronic inflammation results in oxidative stress,³⁷ necrosis and compensatory cell division. Rapid cell division gives rise to an even greater need for DNA repair with its attendant potential for replication errors, which, particularly when they occur at key regulatory sites, such as p53, increase the risk of mutagenesis.¹ Finally, inflammation involves the release of cytokines and growth factors. Although how the specific biology of chronic inflammation relates to cancer is unknown, increasing agreement

focuses on the balance between the two broad arms of the immune response: cell mediated immunity (CMI) and humoural immunity (HI).2 CMI is associated with CD4+ lymphocytes that produce non-specific pro-inflammatory cytokines (also called T-helper 1 (Th-1) cytokines). HI is associated with CD4+ lymphocytes that produce immune suppressive cytokines (also called T-helper 2 (Th-2) cytokines). Th-1 cytokines include interferon- γ , TNF- α , and IL-1 α and IL-1 β . Th-2 cytokines include IL-4, IL-10 and TNF-β. Some cytokines, such as IL-6, have been classified with Th-1 cytokines by some authors and with Th-2 by others.3,38 Pro-inflammatory cytokines are expressed in the presence of infections and inflammants. They upregulate cyclo-oxygenase (COX)-2, the enzyme that catalyses the synthesis of prostaglandins. Prostaglandins, in turn, decrease cell differentiation, inhibit apoptosis³⁹ increase tumour cell proliferation⁴⁰ and induce angiogenesis through growth factors and matrix metalloproteases. 41 In particular, cytokines such as IL-6 and VEGF are pro-angiogenic, supporting the growth of new blood vessels required for breast tumour growth. 42,43 All of these processes increase the risk for promulgating DNA mutations and tumour spread. Th-2 cytokines, on the other hand, inhibit Th-1 cytokines and COX-2 expression.^{2,44} Certain chronic inflammatory conditions associated with tumour development have been associated with an abundance of Th-2 and a paucity of Th-1 cytokines; however, an imbalance of either Th-1 or Th-2 cytokines, prostaglandins, and COX-2 may play a role in carcinogenesis.

4.1. Inflammation and breast cancer

Indirect epidemiological evidence of a link between inflammation and breast cancer comes from studies showing protection by non-steroidal anti-inflammatory drugs (NSAIDs) and risk from antibiotics. NSAIDs inhibit breast tumour cell growth in vitro and in animal models. 45-49 Both case-control and cohort studies suggest that NSAIDs reduce breast cancer risk.50-68 In the Women's Health Initiative Observational Study, a cohort of more than 80,000 women, regular NSAID use for 5+ years was associated with a 19% decrease in the relative risk (RR) of breast cancer (RR 0.81, 95% CI 0.68-0.97). 51 Results from the Long Island Breast Cancer Prevention Project suggest that this protection may be limited to post-menopausal women (ibuprofen OR 1.08 versus 0.74 for pre-menopausal versus post-menopausal women; aspirin OR 0.89 versus 0.73, pre-menopausal versus post-menopausal women).⁵² In contrast, the use of prescribed antibiotics may elevate the risk for breast cancer. In a case control study of more than 10,000 women, Velicer and colleagues demonstrated that antibiotic use was more common in breast cancer cases and particularly in cases who had used antibiotics for longer duration.⁶⁹ Although there are many potential explanations for this unique finding, one is that the link might be accounted for by antibiotic treatment of chronic inflammatory conditions.⁷⁰

More direct evidence that inflammation plays a role in breast carcinogenesis follows. Within the normal human breast, the Th-1 cytokine IL-1 controls the expression of growth factors⁷¹ and in breast tumours, IL-1 production results in an autocrine and paracrine induction of pro-meta-

static genes⁷² including proteolytic enzymes involved in tumour growth and metastasis, such as matrix metalloproteinase (MMP)-1,2,3 and 9.⁷³ Both IL-6 and its soluble receptor are expressed in breast tumours74 and IL-6 levels are higher in women with breast cancer. 75 IL-8 and its receptors are over-expressed in breast cancer and through regulation of MMP-2 activity, IL-8 may also play a role in the invasiveness of breast cancer.⁷⁶ Finally, IL-10, a Th-2 immune depressive cytokine, inhibiting the production of interferon- γ and TNF- α , in the human breast, appears to be involved in mammary involution.⁷⁷ Although these data suggest that breast cancer involves inflammation, it does not imply that immune reactivity induces breast cancer. The latter is implied by recent findings that the TNF- α – 308A polymorphism, associated with increased TNF-α production^{78–83} is more frequent in patients with breast cancer⁸⁴ and that the TGF-beta1 genotype is associated with an increased risk for breast cancer. 85 Thus, Th-1 upregulation may actually predispose to breast cancer.

4.2. Inflammation and ovarian cancer

We have previously suggested that inflammation may also influence the development of ovarian cancer. Bearing children, using oral contraceptives and breastfeeding, which have been shown consistently protect against ovarian cancer, suppress ovulation,86 a process that involves inflammatory mediators and involves repair of the epithelial wound created by extrusion of the ovum.87,88 Other established epidemiological risk factors for ovarian cancer, many of which cannot be explained by hormones alone or by ovulation suppression, are also associated with inflammation. These include using talcum powder, a crystal with an asbestoslike structure,6,89-95 pelvic inflammatory disease,96,97 and hyperthyroidism.6 As with breast cancer, the use of antiinflammatory agents, such as aspirin and NSAIDS, which inhibit mitotic activity in ovarian cancer cell lines98 appear to protect against the disease.99-103

Th-1 cytokines including TNF- α , IL-6, and IL-1 α and β are elevated in epithelial ovarian tumours, in ascitic fluid around tumours, or in serum of women with ovarian cancer; the level of TNF- α expression correlates with tumour grade. ^{104–107} IL-8 is expressed in the normal human ovary where it is also regulated by IL-1¹⁰⁸ IL-8 may inhibit ovarian epithelial apoptosis, predisposing to ovarian cancer. 109-111 Finally, a polarised IL-10 response, albeit a Th-2 cytokine, appears to be produced by ovarian tumours. 112-114 A panel of serum markers, including IL-6, IL-8 and VEGF and reduced concentrations of EGF and IL-12p40 strongly distinguished women with early stage ovarian cancer from controls. 115 With regard to predisposition to ovarian cancer, the IL-1RN gene has a penta-allelic 86-bp tandem repeat (VNTR) in intron 2, of which the less common allele 2 has generally been associated with a reduction in expression116-118 and an elevation in auto-immune conditions. In one study, the VNTR*2 allele was associated with a 2.7-fold elevated risk for ovarian cancer. 119 Moreover, in a nude mouse model, treatment of ascitic ovarian cancer with TNF- α promoted solid tumour nodule formation, and overexpression of TNF- α conferred invasive properties on some tumour cell lines.3,120

5. Hormones in breast and ovarian cancer

Oestrogens, progesterone and androgens are all believed to contribute to breast cancer development, especially in postmenopausal women^{121–126} Early menarche, late menopause and length of reproductive life, factors consistent with prolonged exposure to endogenous oestrogens, have consistently been associated with an increased risk of breast cancer in post-menopausal women. 127 In older post-menopausal women, serum concentrations of oestradiol predict the risk of breast cancer: in a recent pooled analysis of nine prospective studies¹²⁸ a single measurement in the highest, compared with the lowest quintile of free oestradiol doubled the risk for developing post-menopausal breast cancer (RR 2.58, 1.76-3.78). Body mass index (BMI), which increases oestrogen levels in post-menopausal women, 129 elevates the risk for breast cancer by about 3% per kg/m². 130 Moreover, bone mineral density, an integrated marker of long-term oestrogen exposure, is positively associated with breast cancer risk. 131,132 Hormone therapy (HT), the primary source of exogenous oestrogen in post-menopausal women, may increase the risk of breast cancer. 133 However, recent data from the Women's Health Initiative randomised clinical trial suggested that this increase is associated only with oestrogen + progestin formulations; women on oestrogen-only formulations do not appear to have an increase in breast cancer incidence (HR 0.77, 95% CI 0.59-1.01). 134 These data, together with previous data showing a modest increase in breast cancer risk with oestrogen alone, but a greater risk for oestrogen plus progestin, 135 suggest that progestins may enhance breast cancer risk, an observation consistent with the fact that maximum mitotic activity in breast tissue coincides with maximum progesterone levels during the luteal phase of the menstrual cycle.136

Elevated serum or urinary androgen levels have also been associated with the development of post-menopausal breast cancer in at least seven cohort studies. 124–126,137–140 Two cohort studies did not show an association, 141,142 but both were small (n = 15 and 39 cases, respectively) and one involved pre-menopausal women. 141,142 A pooled analysis of cohort studies estimated that the relative risk of breast cancer in women with testosterone in the top as compared with the bottom quintile was 2.2 (95% CI 1.6-3.1) and that the dose-response relationship between testosterone and breast cancer risk was statistically significant. 143 Two of the cohort studies showed a correlation with free testosterone, the measure of bioavailable testosterone. Moreover, several case-control studies have shown that women with breast cancer have elevated levels of testosterone. 144,145 Thus, consistently, observational data have linked plasma testosterone levels to breast cancer risk.

Ovarian cancer risk may also be influenced by sex steroid hormones, specifically unopposed oestrogens and androgens. Pregnancy and oral contraceptive use, both markedly protective against ovarian cancer^{89,146–165} are associated with a reduction in lifetime exposure to unopposed oestrogens (since both involve oestrogen + progesterone)^{166,167} whereas, early menarche and late menopause increase ovarian cancer risk. Moreover, progestin-only oral contraceptives, which do not totally suppress ovulation, are as protective against ovarian cancer as oestrogen-progestin formulas.¹⁶⁸ Finally, HT in-

creases the risk of ovarian cancer^{169,170} and in one study, this risk was limited to formulations involving oestrogen alone or sequential oestrogen + progestin (wherein most of the month involves ingestion of oestrogen alone).¹⁷¹

Elevated androgen levels may also play a role in ovarian cancer. Androgens are produced by ovarian theca cells, are present in follicular fluid, and are the principal sex steroid of growing ovarian follicles¹⁷² Moreover, the post-menopausal ovary is androgenic 173 and androgen receptors are found in normal ovaries, 174 further supporting the activity of androgens within the organ. In addition, oral contraceptives suppress ovarian testosterone production by 35-70%. 175-179 A prospective study¹⁸⁰ found significantly higher levels of androstenedione in the serum of cases compared with controls, and polycystic ovarian syndrome (PCOS), a condition that elevates androgens 181-183 is associated with ovarian cancer. 184 Finally, androgenic agents used to treat endometriosis may increase ovarian cancer risk¹⁸⁵ It is unclear whether androgen directly impacts ovarian cancer risk, or whether the excess risk involves androgen conversion to oestrogen via aromatase.

Thus, sex hormones appear to play a role in the development of breast and ovarian cancers, although the specific hormone–disease relationships differ by cancer type. In particular, although androgens and oestrogen appear to be involved in both cancers, progesterone may be mitogenic in the breast and protective in the ovary.

6. Endometriosis: interaction between inflammation and hormones

Altered immune function in women with endometriosis is ubiquitous, may favour the growth and progression of endometriosis, and upregulates local oestrogen, a major driver of endometriosis. Sites of endometriosis are surrounded by peritoneal inflammatory cells, 186-188 but rather than clearing the ectopic endometrium, these cells (e.g., natural killer (NK) cells) express reduced cytotoxicity. 189-191 Resection of endometriotic foci increased the percentage of moderately differentiated NK cells, suggesting that endometriosis disturbs differentiation of NK cells. 192 Peripheral and peritoneal T-cells and macrophages are also increased and activated in women affected by endometriosis, 193-197 yet they do not clear endometriotic foci, presumably because of aberrant cytokine and growth factor production. ¹⁹⁸ For example, high levels of peritoneal TGF-β, typically found in the peritoneal fluid surrounding implants, inhibits NK activity around endometriosis. 199 Moreover, with the exception of high levels of TNF- α , Th-2 cytokines dominate the peritoneal environment and Th-1 cytokines dominate the internal endometriosis environment. 186,200–203 Peritoneal macrophages also produce MMP-9 and VEGF, vascular permeability and angiogenesis factors that may enhance the vascular support for ectopic endometrium. 204-206

B cell function and antibody production also appear to be abnormal in endometriosis. Indeed, endometriosis has several of the characteristics of an auto-immune disease. ^{188,207} In a recent study of more than 3600 volunteers with surgically verified endometriosis, rates of reported Sjogren's syndrome were 24 times that expected in the general population, rates of systemic lupus erythematosus were 20 times higher than

expected, and thyroiditis was six times higher than expected.²⁰⁸ Biological features characteristic of auto-immune disease that are found in endometriosis include: T and B lymphocyte aberrancies, multiple organ tissue damage, polyactivation, lymphocyte and apoptosis. 187,207,209-211 These aberrancies may help ectopic endometrium to escape destruction. Treatments which shrink foci, such as Danazol or GnRH agonists, return depressed levels of apoptosis and elevated levels of auto-antibodies to normal. 212-214 In combination, these observations suggest that 'successful' endometriosis uses the immune system to favour its own growth and invasion by manufacturing cytokines and growth factors as well as potentially co-opting host immune defences.

Endometriosis appears to be a condition stimulated by oestrogens and inhibited by progesterone. Women are most likely to acquire the condition in their reproductive years. Many markers of excess unopposed oestrogen tend to elevate the occurrence of endometriosis. 215 Notably, these risk factors overlap with those for ovarian cancer and breast cancer. They include early menarche^{216–220} short or long menstrual cy $cle^{216-218,221,222}$ and tall stature. 184 Pregnancy $^{223-225}$ and oral contraceptive use^{216,218,222,226} may reduce endometriosis risk. These relationships are difficult to deconstruct, however, because oral contraceptives are a staple of treatment for endometriosis, and endometriosis causes infertility. More direct evidence relating oestrogens to endometriosis comes from an autopsy study of 399 rhesus monkeys, 227 20% of whom were serendipitously found at death to have endometriosis. Exposure to oestrogen implants was associated with a 9-fold excess of endometriosis.

The amount of local oestrogen to which endometriotic tissue is exposed probably exceeds systemic levels. This is because ectopic endometrium converts androstenedione to oestrone and oestradiol. 228 In ectopic endometrium, steroidogenic factor-1 (SF-1) a transcription factor capable of stimulating aromatase production, binds more avidly to the aromatase promoter site than does competing binding protein (COUP-TF) a factor that inhibits transcription. Therefore, the aromatase gene is preferentially expressed. In contrast, normal, eutopic endometrium does not express SF-1 and therefore does not produce aromatase. 229 An excess of endogenous androgens might then be expected to drive endometriotic progression. Yet synthetic androgens are used to treat endometriosis. The explanation for this apparent inconsistency may be that exogenous androgens downregulate pituitary gonadotrophins and thus oestrogen production. The overall effect of exogenous androgens may thus be to reduce oestrogenic stimulation of endometriotic foci.

Further autocrine exposure to oestrogen is accomplished by the synthesis of oestrone to oestradiol by ectopic endometrium. Oestradiol is substantially more potent than oestrone. Its conversion from oestrone is accomplished by the enzyme 17 β -HSD type 1. 230,231 The converse situation: conversion to oestrone from oestradiol, is accomplished by transcription of a separate gene, producing the enzyme 17 β -HSD type 2. $^{232-234}$ In ectopic endometrium, expression of 17 β -HSD type 2 is absent. 235 Thus, ectopic endometrium synthesises oestradiol, the more potent form of oestrogen, without having any mechanism to convert it back to oest-

rone. In normal endometrium, glandular cells express large amounts of 17β -HSD type 2, induced by progesterone. Although endometriotic foci are resistant to the anti-oestrogenic effects of progesterone, the fact that progestins are a standard and effective treatment modality for endometriosis suggests that this resistance can be overcome.

A central activity of oestradiol in enlarging endometriotic foci appears to be that it promotes MMP expression. 236-239 Moreover, oestrogen induces COX-2, which gives rise to increased concentrations of prostaglandin (PGE2). 229 Furthermore, PGE2 stimulates aromatase, the enzyme that converts androstenedione to oestrone and oestradiol. 240 This interplay between PGE₂ and aromatase creates a positive feedback loop within ectopic endometrium, resulting in local elevations in pro-inflammatory prostaglandins and gens^{228,240,241} Thus, endometriotic foci enrich the oestrogen in their local environment, both through conversion of systemic androstenedione to oestrone and oestrone to oestradiol and through aromatase, 17\beta -HSD type 1, but also through stimulation of PGE2. That is, in endometriosis, a relatively well-understood inflammatory-hormonal escalation promotes tumour development.

7. Sex hormones and inflammation in relation to breast and ovary cancer

Inflammatory mediators have emerged as important regulators of oestrogen synthesis in breast tissue as well. IL-6 levels increase the activity of aromatase in breast and adipose tissue, thereby directly increasing systemic and local breast oestrogen levels. Within the breast, TNF- α can also act alone or synergistically with IL-6 to enhance the activities of aromatase teach the E2DH and oestrone sulphatase three enzymes involved in the production of oestradiol from androstenedione, oestrone and oestrone sulphate, respectively. IL-1 α levels inversely correlate with local sex steroid receptor expression suggesting an interplay between the cytokine and steroid hormones within breast cancer cells. Finally, IL-8 influences the activity of 17-HSD, enhancing oestrogenic activity.

Conversely, sex steroid hormones directly affect cytokines. Series For example, oestrogen receptor complexes can bind directly to the IL-6 promoter region, sex resulting in decreased IL-6 promoter activity. In addition, expression of many cytokines and growth factors is menstrual cycle dependent and hormone therapy (HT) may impact the expression of several cytokines, including IL-2, IL-6, TNF- α and IFN- $\gamma^{254-257}$ 2-MeOE2, a metabolite of oestradiol associated with reduced risk of breast cancer, sex inhibits the production and actions of both IL-6 and TNF- α .

8. Cytokines and hormones interact in the ovary

There is also mounting evidence of hormone–cytokine interactions within the ovary. For example, ovulation involves increased production of TNF- α , IL-1 β , IL-6 and COX-2. Indeed, the anti-inflammatory NSAIDs inhibit human ovulation. ²⁶³ Ovarian cancers are thought to arise within inclusion cysts, epithelium that folds into the stroma as surface cells

are entrapped in the ovarian wound created during ovulation. This brings epithelium into close proximity with stromal inflammatory mediators involved in ovulation. 1,88,261,264 Steroid hormones in the ovarian stroma and epithelium modulate both Th-1 cytokines and TGF- β expression. 265,266 Finally, cytokines and growth factors including IL-1, IFN- γ , TNF- α and IGF-1 directly stimulate ovarian steroidogenesis $^{267-269}$ and may play a role in both ovulation and oestrogen-induced tumour growth. 251

Hence, as in endometriosis, in both the ovary and breast, there appears to be an interaction between sex steroid hormones and cytokines/growth factors. It can be readily postulated that the mutual upregulation of these systems may promote carcinogenesis in both reproductive organs.

9. Summary

Endometriosis is associated with both ovarian and breast cancers. The relationship with ovarian cancer can be understood as a local process of malignant transformation, whereas the relationship with breast cancer cannot. Sex steroid hormones and inflammation have been implicated in the pathogenesis of each of endometriosis, ovarian cancer and breast cancer. A possible explanation for the association between endometriosis and these reproductive cancers is one of local and systemic enhancement of aberrant inflammatory and hormonal mediators. In susceptible tissues, i.e., ones wherein inflammatory–hormonal interactions are most influential, the impact of endometriosis may be to promote a feed-forward loop that favours cancer progression.

Several clinical considerations arise from an emerging understanding of how endometriosis might mediate cancers. First, endometriosis may well be considered a risk factor for ovarian and breast cancers, triggering increasingly intensive surveillance and the possible use of chemoprevention in affected women. Secondly, treatments for endometriosis may best take into consideration the impact on long-term cancer risk. For example, increasing circulating steroid hormones may be harmful long-term, whereas modulation of inflammatory balance may be beneficial. Thirdly, the evolving basic science that characterises how hormones and inflammatory mediators interact within endometriotic implants may have relevance beyond endometriosis. Other cancers, such as prostate, colon and endometrial, may involve hormone-inflammatory interactions. Finally, the notion that a nidus of inflammation in one tissue could have effects on a distant tissue may have extensive implications on other inflammationcancer relationships.

Conflict of interest statement

None declared.

REFERENCES

 Ames BN, Gold LS, Willet WC. The causes and prevention of cancer. Proc Natl Acad Sci USA 1995;92:5258–65.

- O'Byrne KJ, Dalgleish AG. Chronic immune activation and inflammation as the cause of malignancy. Br J Cancer 2001;85:473–83.
- 3. Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet* 2001;357:539–45.
- Edwards JG, Abrams KR, Leverment JN, Spyt TJ, Waller DA, O'Byrne KJ. Prognostic factors for malignant mesothelioma in 142 patients: validation of CALGB and EORTC prognostic scoring systems. Thorax 2000;55:731–5.
- 5. Monson RR, Yen S, MacMahon B. Chronic mastitis and carcinoma of the breast. *Lancet* 1976;2:224–6.
- Ness RB, Grisso JA, Cottreau C, et al. Factors related to inflammation of the ovarian epithelium and risk of ovarian cancer. Epidemiology 2000;11:111–7.
- Ness RB, Cottreau C. Possible role of ovarian epithelial inflammation in ovarian cancer. J Natl Cancer Inst 1999;91:1459–67.
- 8. Cibere J, Sibley J, Haga M. Rheumatoid arthritis and the risk of malignancy. Arthritis Rheum 1997;40:1580–6.
- Brinton LA, Gridley G, Persson I, Baron J, Bergqvist A. Cancer risk after a hospital discharge diagnosis of endometriosis. Am J Obstet Gynecol 1997;176:572–9.
- Brinton LA, Lamb EJ, Moghissi KS, et al. Ovarian cancer risk associated with varying causes of infertility. Fertil Steril 2004;82:405–14.
- Duczman L, Ballweg ML. Endometriosis and cancer: what is the connection?, vol 20. Milwaukee (WIS): Endometriosis Association Newsletter; 1999.
- Ness RB, Cramer DW, Goodman MT, et al. Infertility, fertility drugs, and ovarian cancer: a pooled analysis of case-control studies. Am J Epidemiol 2002;155:217–24.
- Cahill DJ, Hull MG. Pituitary-ovarian dysfunction and endometriosis. Hum Reprod Update 2000;6:56–66.
- Modugno F, Ness RB, Allen GO, Schildkraut JM, Davis FG, Goodman MT. Oral contraceptive use, reproductive history, and risk of epithelial ovarian cancer in women with and without endometriosis. Am J Obstet Gynecol 2004;191:733–40.
- Schairer C, Persson I, Falkeborn M, Naessen T, Troisi R, Brinton LA. Breast cancer risk associated with gynecologic surgery and indications for such surgery. Int J Cancer 1997;70:150–4.
- Rossing MA, Daling JR, Weiss NS, Moore DE, Self SG. Ovarian tumors in a cohort of infertile women. New Engl J Med 1994;331:771–6.
- Ron E, Lunenfeld B, Menczer J, et al. Cancer incidence in a cohort of infertile women. Am J Epidemiol 1987;125:780–90.
- Venn A, Watson L, Lumley J, Giles G, King C, Healy D. Breast and ovarian cancer incidence after infertility and in vitro fertilization. *Lancet* 1995;346:995–1000.
- 19. Olson JE, Cerhan JR, Janney CA, Anderson KE, Vachon CM, Sellers TA. Postmenopausal cancer risk after self-reported endometriosis diagnosis in the Iowa women's health study. *Cancer* 2002;**94**:1612–8.
- Chang CK, Chen P, Leu FJ, Lou SM. Florid polypoid endometriosis exacerbated by tamoxifen therapy in breast cancer. Obstet Gynecol 2003;102:1127–30.
- 21. Bese T, Simsek Y, Bese N, Ilvan S, Arvas M. Extensive pelvic endometriosis with malignant change in tamoxifen-treated postmenopausal women. *Int J Gynecol Cancer* 2003;**13**:376–80.
- 22. Heaps JM, Nieberg RK, Berek JS. Malignant neoplasms arising in endometriosis. Obstet Gynecol 1990;75:1023–8.
- Sainz de la Cuesta R, Eichhorn JH, Rice LW, Fuller Jr AF, Nikrui BA, Goff BA. Histologic transformation of benign endometriosis to early epithelial ovarian cancer. Gynecol Oncol 1996;60:238–44.
- Vercellini P, Parazzini F, Bolis G, et al. Endometriosis and ovarian cancer. Am J Obstet Gynecol 1993;169:181–2.

- 25. LaGrenade A, Silverberg SG. Ovarian tumors associated with atypical endometriosis. *Hum Pathol* 1988;19:1080–4.
- Moll UM, Chumas JC, Chalas E, Mann WJ. Ovarian carcinoma arising in atypical endometriosis. Obstet Gynecol 1990;75:537–9.
- 27. Jimbo H, Yoshikawa H, Onda T, Yasugi T, Sakamoto A, Taketani Y. Prevalence of ovarian endometriosis in epithelial ovarian cancer. Int J Gynaecol Obstet 1997;59:245–50.
- DePriest PD, Banks ER, Powell DE, et al. Endometrioid carcinoma of the ovary and endometriosis: the association in postmenopausal women. Gynecol Oncol 1992;47:71–5.
- Stern RC, Dash R, Bentley RC, Snyder MJ, Haney AF, Robboy SJ. Malignancy in endometriosis: frequency and comparison of ovarian and extraovarian types. Int J Gynecol Pathol 2001;20:133–9.
- Mostoufizadeh M, Scully RE. Malignant tumors arising in endometriosis. Clin Obstet Gynecol 1980;23:951–63.
- Sampson JA. Endometrial carcinoma of the ovary arising in endometrial tissue in that organ. Arch Surg 1925;10:1–72.
- Hitti IF, Glasberg SS, Lubicz S. Clear cell carcinoma arising in extraovarian endometriosis: report of three cases and review of the literature. Gynecol Oncol 1990;39:314–20.
- Erzen M, Kovacic J. Relationship between endometriosis and ovarian cancer. Eur J Gynaecol Oncol 1998;19:553–5.
- 34. Friedlander ML. Prognostic factors in ovarian cancer. Semin Oncol 1998;25:305–14.
- Fukunaga M, Nomura K, Ishikawa E, Ushigome S. Ovarian atypical endometriosis: its close association with malignant epithelial tumours. Histopathology 1997;30:249–55.
- Yoshikawa H, Jimbo H, Okada S, et al. Prevalence of endometriosis in ovarian cancer. Gynecol Obstet Invest 2000;50:11–7.
- Christensen S, Hagen TM, Shigenaga MK, Ames BN. Chronic inflammation, mutation and cancerMicrobes and malignancy: infection as a cause of human cancers. New York: Oxford University Press; 1999.
- 38. Mosmann TR, Coffman RL. TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. Annu Rev Immunol 1989;7:145–73.
- Sheng H, Shao J, Morrow JD, Beauchamp RD, DuBois RN. Modulation of apoptosis and Bcl-2 expression by prostaglandin E2 in human colon cancer cells. Cancer Res 1998;58:362-6.
- Qiao L, Kozoni V, Tsioulias GJ, et al. Selected eicosanoids increase the proliferation rate of human colon carcinoma cell lines and mouse colonocytes in vivo. Biochimica et Biophysica Acta 1995;1258:215–23.
- 41. Taketo MM. Cyclooxygenase nfl-2 inhibitors in tumorigenesis (part II). J Natl Cancer Inst 1998;90:1609–20.
- Cross MJ, Claesson-Welsh L. FGF and VEGF function in angiogenesis: signaling pathways, biological responses and therapeutic inhibition. Trends Pharmacol Sci 2001;22:201–7.
- 43. Folkman J. Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nat Med* 1995;1:27–31.
- 44. Clerici M, Shearer GM, Clerici E. Cytokine dysregulation in invasive cervical carcinoma and other human neoplasias: time to consider the TH1/TH2 paradigm. *J Natl Cancer Inst* 1998;**90**:261–3.
- 45. Alshafie GA, Harris RE, Robertson FM, Parrett ML, Ross M, Abou-Issa H. Comparative chemopreventive activity of ibuprofen and N-(4-hydroxyphenyl) retinamide against the development and growth of rat mammary adenocarcinomas. Anticancer Res 1999;19:3031–6.
- Ip MM, Mazzer C, Watson D, Ip C. The effect of eicosanoid synthesis inhibitors on DMBA-induced rat mammary carcinogenesis. Proc Am Assoc Cancer Res 1989;30:182.

- 47. Joarder FS, Abou-Issa H, Robertson FM, Parrett ML, Alshafie G, Harris RE. Growth arrest of DMBA-induced mammary carcinogenesis with ibuprofen treatment in female Sprague–Dawley rats. Proc Am Assoc Cancer Res 1997;38:370.
- 48. Lee PP, Ip MM. Regulation of proliferation of rat mammary tumor cells by inhibitors of cyclooxygenase and lipoxygenase. Prostaglandins Leukot Essent Fatty Acids 1992;45:21–31.
- 49. Steele VE, Moon RC, Lubet RA, et al. Preclinical efficacy evaluation of potential chemopreventive agents in animal carcinogenesis models: methods and results from the NCI chemoprevention drug development program. *J Cell Biochem Suppl* 1994;20:32–54.
- Thun MJ, Namboodiri MM, Calle EE, Flanders WD, Heat Jr CW. Aspirin use and risk of fatal cancer. Cancer Res 1993:53:1322-7.
- 51. Harris RE, Chlebowski RT, Jackson RD, et al. Breast cancer and nonsteroidal anti-inflammatory drugs: prospective results from the women's health initiative. *Cancer Res* 2003;63:6096–101.
- 52. Terry MB, Gammon MD, Zhang FF, et al. Association of frequency and duration of aspirin use and hormone receptor status with breast cancer risk. *J Am Med Assoc* 2004;291:2433–40.
- 53. Coogan PF, Rao SR, Rosenberg L, et al. The relationship of nonsteroidal anti-inflammatory drug use to the risk of breast cancer. *Prev Med* 1999;29:72–6.
- 54. Cotterchio M, Kreiger N, Sloan M, Steingart A. Nonsteroidal anti-inflammatory drug use and breast cancer risk. Cancer Epidemiol Biomarkers Prev 2001;10:1213–7.
- 55. Egan KM, Stampfer MJ, Giovannucci E, Rosner BA, Colditz GA. Prospective study of regular aspirin use and the risk of breast cancer. J Natl Cancer Inst 1996;88:988–93.
- 56. Gridley G, McLaughlin JK, Ekbom A, et al. Incidence of cancer among patients with rheumatoid arthritis. *J Natl Cancer Inst* 1993;**85**:307–11.
- Harris RE, Namboodiri KK, Farrar WB. Epidemiologic study of non-steroidal anti-inflammatory drugs and breast cancer. Oncol Rep 1995;2:591–2.
- Harris RE, Namboodiri KK, Farrar WB. Nonsteroidal anti-inflammatory drugs and breast cancer. Epidemiology 1996;7:203–5.
- Harris RE, Kasbari S, Farrar WB. Prospective study of nonsteroidal anti-inflammatory drugs and breast cancer. Oncol Rep 1999;6:71–3.
- Johnson TJ, Anderson KI, Lazovich D, Folsom AR. Association of aspirin and other nonsteroidal anti-inflammatory drug use with incidence of postmenopausal breast cancer. Proc Am Assoc Cancer Res 2001;42:763.
- 61. Khuder SA, Mutgi AB. Breast cancer and NSAID use: a meta-analysis. Br J Cancer 2001;84:1188–92.
- 62. Langman MJ, Cheng KK, Gilman EA, Lancashire RJ. Effect of anti-inflammatory drugs on overall risk of common cancer: case-control study in general practice research database. Br Med J 2000;320:1642–6.
- 63. Neugut AI, Rosenberg DJ, Ahsan H, et al. Association between coronary heart disease and cancers of the breast, prostate, and colon. Cancer Epidemiol Biomarkers Prev 1998;7:869–73.
- 64. Paganini-Hill A, Chao A, Ross RK, Henderson BE. Aspirin use and chronic diseases: a cohort study of the elderly. Br Med J 1989:299:1247–50.
- 65. Rosenberg L. Nonsteroidal anti-inflammatory drugs and cancer. *Prev Med* 1995;**24**:107–9.
- 66. Schreinemachers DM, Everson RB. Aspirin use and lung, colon, and breast cancer incidence in a prospective study. Epidemiology 1994;5:138–46.

- 67. Sharpe CR, Collet JP, McNutt M, Belzile E, Boivin JF, Hanley JA. Nested case-control study of the effects of non-steroidal anti-inflammatory drugs on breast cancer risk and stage. Br J Cancer 2000;83:112–20.
- 68. Harris RE, Namboodiri K, Stellman SD, Wynder EL. Breast cancer and NSAID use: heterogeneity of effect in a case-control study. *Prev Med* 1995;24:119–20.
- Velicer CM, Heckbert SR, Lampe JW, Potter JD, Robertson CA, Taplin SH. Antibiotic use in relation to the risk of breast cancer. J Am Med Assoc 2004;291:827–35.
- 70. Ness RB, Cauley JA. Antibiotics and breast cancer what's the meaning of this? J Am Med Assoc 2004;291:880–1.
- 71. Palmieri C, Roberts-Clark D, Assadi-Sabet A, et al. Fibroblast growth factor 7, secreted by breast fibroblasts, is an interleukin-1beta-induced paracrine growth factor for human breast cells. *J Endocrinol* 2003;177:65–81.
- 72. Nozaki S, Sledge Jr GW, Nakshatri H. Cancer cell-derived interleukin 1alpha contributes to autocrine and paracrine induction of pro-metastatic genes in breast cancer. *Biochem Biophys Res Commun* 2000;275:60–2.
- 73. Rawdanowicz TJ, Hampton AL, Nagase H, Woolley DE, Salamonsen LA. Matrix metalloproteinase production by cultured human endometrial stromal cells: identification of interstitial collagenase, gelatinase-A, gelatinase-B, and stromelysin-1 and their differential regulation by interleukin-1 alpha and tumor necrosis factor-alpha. *J Clin Endocrinol Metab* 1994;79:530–6.
- Knupfer H, Schmidt R, Stanitz D, Brauckhoff M, Schonfelder R, Preiss R. CYP2C and IL-6 expression in breast cancer. Breast 2004;13:28–34.
- 75. Kozlowski L, Zakrzewska I, Tokajuk P, Wojtukiewicz MZ. Concentration of interleukin-6 (IL-6), interleukin-8 (IL-8) and interleukin-10 (IL-10) in blood serum of breast cancer patients. Rocz Akad Med Bialymst 2003;48:82–4.
- Lin Y, Huang R, Chen L, et al. Identification of interleukin-8 as estrogen receptor-regulated factor involved in breast cancer invasion and angiogenesis by protein arrays. Int J Cancer 2004;109:507–15.
- 77. Sohn BH, Moon HB, Kim TY, et al. Interleukin-10 upregulates tumour-necrosis-factor-alpha-related apoptosis-inducing ligand (TRAIL) gene expression in mammary epithelial cells at the involution stage. Biochem J 2001;360:31–8.
- Galbraith GM, Steed RB, Sanders JJ, Pandey JP. Tumor necrosis factor alpha production by oral leukocytes: influence of tumor necrosis factor genotype. J Periodontol 1998:69:428–33.
- Grove J, Daly AK, Bassendine MF, Day CP. Association of a tumor necrosis factor promoter polymorphism with susceptibility to alcoholic steatohepatitis. Hepatology 1997;26:143–6.
- Huang DR, Pirskanen R, Matell G, Lefvert AK. Tumour necrosis factor-alpha polymorphism and secretion in myasthenia gravis. J Neuroimmunol 1999;94:165–71.
- 81. Kroeger KM, Steer JH, Joyce DA, Abraham LJ. Effects of stimulus and cell type on the expression of the –308 tumour necrosis factor promoter polymorphism. Cytokine 2000;12:110–9.
- 82. Maurer M, Kruse N, Giess R, Kyriallis K, Toyka KV, Rieckmann P. Gene polymorphism at position –308 of the tumor necrosis factor alpha promotor is not associated with disease progression in multiple sclerosis patients. *J Neurol* 1999;246:949–54.
- Obayashi H, Nakamura N, Fukui M, et al. Influence of TNF microsatellite polymorphisms (TNFa) on age-at-onset of insulin-dependent diabetes mellitus. Hum Immunol 1999;60:974–8.
- 84. Chouchane L, Ahmed SB, Baccouche S, Remadi S. Polymorphism in the tumor necrosis factor-alpha promotor

- region and in the heat shock protein 70 genes associated with malignant tumors. *Cancer* 1997;**80**:1489–96.
- 85. Ziv E, Cauley J, Morin PA, Saiz R, Browner WS. Association between T29 → C polymorphism in the transforming growth factor beta1 gene and breast cancer among elderly women: the study of osteoporotic fractures. J Am Med Assoc 2001;285:2859–63.
- 86. Fathalla MF. Incessant ovulation a factor in ovarian neoplasia? *Lancet* 1971;2:163.
- Murdoch WJ. Ovarian surface epithelium, ovulation and carcinogenesis. Biol Rev Camb Philos Soc 1996;71:529–43.
- 88. Auersperg N, Maines-Bandiera SL, Dyck HG. Ovarian carcinogenesis and the biology of ovarian surface epithelium. *J Cell Physiol* 1997;173:261–5.
- 89. Booth M, Beral V, Smith P. Risk factors for ovarian cancer: a case-control study. Br J Cancer 1989;60:592–8.
- Hartge P, Hoover R, Lesher LP, McGowan L. Talc and ovarian cancer. J Am Med Assoc 1983;250:1844.
- 91. Cook LS, Kamb ML, Weiss NS. Perineal powder exposure and the risk of ovarian cancer. *Am J Epidemiol* 1997;**145**:459–65.
- Cramer DW, Welch WR, Scully RE, Wojciechowski CA.
 Ovarian cancer and talc: a case-control study. Cancer 1982:50:372-6.
- 93. Harlow BL, Cramer DW, Bell DA, Welch WR. Perineal exposure to talc and ovarian cancer risk. Obstet Gynecol 1992;80:19–26.
- 94. Whittemore AS, Wu ML, Paffenbarger Jr RS. Personal and environmental characteristics related to epithelial ovarian cancer. II. Exposures to talcum powder, tobacco, alcohol, and coffee. Am J Epidemiol 1988;128:1228–40.
- 95. Tzonou A, Polychronopoulou A, Hsieh CC, Rebelakos A, Karakatsani A, Trichopoulos D. Hair dyes, analgesics, tranquilizers and perineal talc application as risk factors for ovarian cancer. *Int J Cancer* 1993;55:408–10.
- 96. Risch HA, Howe GR. Pelvic inflammatory disease and the risk of epithelial ovarian cancer. *Cancer Epidemiol Biomarkers Prev* 1995;4:447–51.
- 97. Ness RB, Goodman MT, Shen C, Brunham RC. Serologic evidence of past infection with Chlamydia trachomatis, in relation to ovarian cancer. *J Infect Des* 2003;**187**: 1147–1152.
- 98. Rodriguez-Burford C, Barnes MN, Oelschlager DK, et al. Effects of nonsteroidal anti-inflammatory agents (NSAIDs) on ovarian carcinoma cell lines: preclinical evaluation of NSAIDs as chemopreventive agents. Clin Cancer Res 2002:8:202-9
- Akhmedkhanov A, Toniolo P, Zeleniuch-Jacquotte A, Kato I, Koenig KL, Shore RE. Aspirin and epithelial ovarian cancer. Prev Med 2001;33:682–7.
- Tzonou A, Day NE, Trichopoulos D, et al. The epidemiology of ovarian cancer in Greece: a case-control study. Eur J Cancer Clin Oncol 1984;20:1045–52.
- 101. Cramer DW, Harlow BL, Titus-Ernstoff L, Bohlke K, Welch ER, Greenberg ER. Over-the-counter analgesics and risk of ovarian cancer. Lancet 1998;351:104–7.
- 102. Cramer DW. Hormonal and dietary factors potential for chemopreventive of ovarian cancer. Pittsburgh, 2002.
- Rosenberg L, Palmer JR, Rao RS, et al. A case-control study of analgesic use and ovarian cancer. Cancer Epidemiol Biomarkers Prev 2000;9:933–7.
- 104. Naylor MS, Stamp GW, Foulkes WD, Eccles D, Balkwill FR. Tumor necrosis factor and its receptors in human ovarian cancer. Potential role in disease progression. *J Clin Invest* 1993;91:2194–206.
- Burke F, Relf M, Negus R, Balkwill F. A cytokine profile of normal and malignant ovary. Cytokine 1996;8:578–85.
- 106. Asschert JG, Vellenga E, Ruiters MH, de Vries EG. Regulation of spontaneous and TNF/IFN-induced IL-6 expression in two

- human ovarian-carcinoma cell lines. *Int J Cancer* 1999:**82**:244–9.
- Punnonen R, Teisala K, Kuoppala T, Bennett B, Punnonen J. Cytokine production profiles in the peritoneal fluids of patients with malignant or benign gynecologic tumors. Cancer 1998:83:788–96.
- 108. Fujii A, Harada T, Yamauchi N, et al. Interleukin-8 gene and protein expression are upregulated by interleukin-1beta in normal human ovarian cells and a granulose tumor cell line. Fertil Steril 2003;79:151–7.
- 109. Toutirais O, Chartier P, Dubois D, et al. Constitutive expression of TGF-beta1, interleukin-6 and interleukin-8 by tumor cells as a major component of immune escape in human ovarian carcinoma. Eur Cytokine Netw 2003;14:246-55.
- 110. Abdollahi T, Robertson NM, Abdollahi A, Litwack G. Identification of interleukin 8 as an inhibitor of tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis in the ovarian carcinoma cell line OVCAR3. Cancer Res 2003;63:4521–6.
- 111. Xu L, Fidler IJ. Interleukin 8: an autocrine growth factor for human ovarian cancer. Oncol Res 2000;12:97–106.
- 112. Gotlieb WH, Abrams JS, Watson JM, Velu TJ, Berek JS, Martinez-Maza O. Presence of interleukin 10 (IL-10) in the ascites of patients with ovarian and other intra-abdominal cancers. Cytokine 1992;4:385–90.
- 113. Loercher AE, Nash MA, Kavanagh JJ, Platscoucas CD, Freedman RS. Identification of an IL-10-producing HLA-DR-negative monocyte subset in the malignant ascites of patients with ovarian carcinoma that inhibits cytokine protein expression and proliferation of autologous T cells. J Immunol 1999;163:6251–60.
- 114. Pisa P, Halapi E, Pisa EK, et al. Selective expression of interleukin 10, interferon gamma, and granulocytemacrophage colony-stimulating factor in ovarian cancer biopsies. Proc Natl Acad Sci USA 1992;89:7708–12.
- 115. Gorelik E, Landsittel DP, Marrangoni AM, et al. Novel serum multianalyte immunobead diagnostic assay, MIDAS, for early detection of ovarian cancer. 2004, in press.
- 116. Danis VA, Millington M, Hyland VJ, Grennan D. Cytokine production by normal human monocytes: inter-subject variation and relationship to an IL-1 receptor antagonist (IL-1Ra) gene polymorphism. Clin Exp Immunol 1995;99:303–10.
- 117. Hurme M, Santtila S. IL-1 receptor antagonist (IL-1Ra) plasma levels are co-ordinately regulated by both IL-1Ra and IL-1beta genes. Eur J Immunol 1998;28:2598–602.
- 118. Tarlow JK, Blakemore AI, Lennard A, et al. Polymorphism in human IL-1 receptor antagonist gene intron 2 is caused by variable numbers of an 86-bp tandem repeat. Hum Genet 1993;91:403–4.
- Sehouli J, Mustea A, Koensgen D, Lichtenegger W.
 Interleukin-1 receptor antagonist gene polymorphism in epithelial ovarian cancer. Cancer Epidemiol Biomarkers Prev 2003;12:1205–8.
- 120. Malik ST, Griffin DB, Fiers W, Balkwill FR. Paradoxical effects of tumour necrosis factor in experimental ovarian cancer. Int J Cancer 1989;44:918–25.
- 121. Moore JW, Clark GM, Hoare SA, et al. Binding of oestradiol to blood proteins and aetiology of breast cancer. *Int J Cancer* 1986;38:625–30.
- 122. Colditz GA. Relationship between estrogen levels, use of hormone replacement therapy, and breast cancer. *J Natl Cancer Inst* 1998;**90**:814–23.
- 123. Toniolo PG, Levitz M, Zeleniuch-Jacquotte A, et al. A prospective study of endogenous estrogens and breast cancer in postmenopausal women. *J Natl Cancer Inst* 1995;87:190–7.

- 124. Dorgan JF, Longcope C, Stephenson Jr HE, et al. Relation of prediagnostic serum estrogen and androgen levels to breast cancer risk. *Cancer Epidemiol Biomarkers Prev* 1996;5:533–9.
- 125. Berrino F, Muti P, Micheli A, et al. Serum sex hormone levels after menopause and subsequent breast cancer. *J Natl Cancer Inst* 1996;88:291–6.
- 126. Hankinson SE, Willett WC, Manson JE, et al. Plasma sex steroid hormone levels and risk of breast cancer in postmenopausal women. J Natl Cancer Inst 1998;90:1292–9.
- 127. Hulka BS, Stark AT. Breast cancer: cause and prevention. *Lancet* 1995;346:883–7.
- 128. Key T, Appleby P, Barnes I, Reeves G. Endogenous Hormones and Breast Cancer Collaborative Group. Endogenous sex hormones and breast cancer in postmenopausal women: reanalysis of nine prospective studies. *JNCI Cancer Spectrum* 2002:94:606–16.
- 129. Hankinson SE, Willett WC, Manson JE, et al. Alcohol, height, and adiposity in relation to estrogen and prolactin levels in postmenopausal women. *J Natl Cancer Inst* 1995;87:1297–302.
- 130. Beral V. Breast cancer and hormone replacement therapy: collaborative reanalysis of data from 51 epidemiological studies of 52,705 women with breast cancer and 108,411 women without breast cancer. Collaborative Group on Hormonal Factors in Breast Cancer. Lancet 1997;350:1047–59.
- 131. Cauley JA, Lucas FL, Kuller LH, Vogt MT, Browner WS, Cummings SR. Bone mineral density and risk of breast cancer in older women: the study of osteoporotic fractures. Study of Osteoporotic Fractures Research Group. J Am Med Assoc 1996;276:1404–8.
- 132. Buist DSM, LaCroix AZ, Barlow WE, White E, Weiss NS. Bone mineral density and breast cancer risk in postmenopausal women. *J Clinical Epidemiology* 2001;54:417–22.
- 133. Chlebowski RT, Hendrix SL, Langer RD, et al. Influence of estrogen plus progestin on breast cancer and mammography in healthy postmenopausal women: the women's health initiative randomized trial. J Am Med Assoc 2003;289:3243–53.
- 134. Anderson GL, Limacher M, Assaf AR, et al. Effects of conjugated equine estrogen in postmenopausal women: the women's health initiative randomized controlled trial. *J Am Med Assoc* 2004;291:1701–12.
- 135. Ross RK, Paganini-Hill A, Wan PC, Pike MC. Effect of hormone replacement therapy on breast cancer risk: estrogen versus estrogen plus progestin. *J Natl Cancer Inst* 2000;92:328–32.
- 136. Pike MC, Spicer DV, Dahmoush L, Press MF. Estrogens, progestogens, normal breast cell proliferation, and breast cancer risk. *Epidemiol Rev* 1993;15:17–35.
- 137. Wang DY, Allen DS, De Stavola BL, et al. Urinary androgens and breast cancer risk: results from a long-term prospective study based in Guernsey. Br J Cancer 2000;82:1577–84.
- 138. Thomas HV, Key TJ, Allen DS, et al. A prospective study of endogenous serum hormone concentrations and breast cancer risk in post-menopausal women on the island of Guernsey. Br J Cancer 1997;76:401–5.
- 139. Zeleniuch-Jacquotte A, Bruning PF, Bonfrer JM, et al. Relation of serum levels of testosterone and dehydroepiandrosterone sulfate to risk of breast cancer in postmenopausal women. Am J Epidemiol 1997;145:1030–8.
- 140. Cauley JA, Lucas FL, Kuller LH, Stone K, Browner W, Cummings SR. Elevated serum estradiol and testosterone concentrations are associated with a high risk for breast cancer. Study of Osteoporotic Fractures Research Group. Ann Intern Med 1999;130:270–7.
- 141. Wysowski DK, Comstock GW, Helsing KJ, Lau HL. Sex hormone levels in serum in relation to the development of breast cancer. Am J Epidemiol 1987;125:791–9.
- 142. Garland CF, Friedlander NJ, Barrett-Connor E, Khaw KT. Sex hormones and postmenopausal breast cancer: a prospective

- study in an adult community. Am J Epidemiol 1992:135:1220–30.
- 143. Lillie EO, Bernstein L, Ursin G. The role of androgens and polymorphisms in the androgen receptor in the epidemiology of breast cancer. Breast Cancer Res 2003;5:164–73.
- 144. Secreto G, Recchione C, Ballerini P, et al. Accumulation of active androgens in breast cyst fluids. Eur J Cancer 1991;27:44–7.
- 145. Lipworth L, Adami HO, Trichopoulos D, Carlstrom K, Mantzoros C. Serum steroid hormone levels, sex hormonebinding globulin, and body mass index in the etiology of postmenopausal breast cancer. Epidemiology 1996;7:96–100.
- 146. Mori M, Harabuchi I, Miyake H, Casagrande JT, Henderson RK, Ross RK. Reproductive, genetic, and dietary risk factors for ovarian cancer. Am J Epidemiol 1988;128:771–7.
- 147. Wynder EL, Dodo H, Barber HR. Epidemiology of cancer of the ovary. *Cancer* 1969;**23**:352–70.
- 148. Voigt LF, Harlow BL, Weiss NS. The influence of age at first birth and parity on ovarian cancer risk. Am J Epidemiol 1986;124:490–1.
- Risch HA, Weiss NS, Lyon JL, Daling JR, Liff JM. Events of reproductive life and the incidence of epithelial ovarian cancer. Am J Epidemiol 1983;117:128–39.
- 150. Nasca PC, Greenwald P, Chorost S, Richart R, Caputo T. An epidemiologic case-control study of ovarian cancer and reproductive factors. *Am J Epidemiol* 1984;119:705–13.
- 151. La Vecchia C, Decarli A, Franceschi S, Regallo M, Tognoni G. Age at first birth and the risk of epithelial ovarian cancer. J Natl Cancer Inst 1984;73:663–6.
- 152. Franceschi S, La Vecchia C, Helmrich SP, Mangioni C, Tognoni G. Risk factors for epithelial ovarian cancer in Italy. Am J Epidemiol 1982;115:714–9.
- Kvale G, Heuch I, Nilssen S, Beral V. Reproductive factors and risk of ovarian cancer: a prospective study. Int J Cancer 1988;42:246–51.
- 154. Mori M, Kiyosawa H, Miyake H. Case-control study of ovarian cancer in Japan. *Cancer* 1984;**53**:2746–52.
- 155. Rosenberg L, Shapiro S, Slone D, et al. Epithelial ovarian cancer and combination oral contraceptives. *J Am Med Assoc* 1982:247:3210–2
- Casagrande JT, Louie EW, Pike MC, Roy S, Ross RK, Henderson BE. 'Incessant ovulation' and ovarian cancer. Lancet 1979;2:170–3.
- 157. Hildreth NG, Kelsey JL, LiVolsi VA, et al. An epidemiologic study of epithelial carcinoma of the ovary. Am J Epidemiol 1981;114:398–405.
- 158. Gwinn ML, Lee NC, Rhodes PH, Layde PM, Rubin GL. Pregnancy, breast feeding, and oral contraceptives and the risk of epithelial ovarian cancer. J Clin Epidemiol 1990;43:559–68.
- 159. McGowan L, Parent L, Lednar W, Norris HJ. The woman at risk for developing ovarian cancer. Gynecol Oncol 1979:7:325–44.
- 160. Newhouse ML, Pearson RM, Fullerton JM, Boesen EA, Shannon HS. A case control study of carcinoma of the ovary. Br J Prev Soc Med 1977;31:148–53.
- Risch HA. Estrogen replacement therapy and risk of epithelial ovarian cancer. Gynecol Oncol 1996;63:254–7.
- 162. Risch HA, Marrett LD, Howe GR. Parity, contraception, infertility, and the risk of epithelial ovarian cancer. Am J Epidemiol 1994;140:585–97.
- 163. Cramer DW, Hutchison GB, Welch WR, Scully RE, Ryan KJ. Determinants of ovarian cancer risk. I. Reproductive experiences and family history. J Natl Cancer Inst 1983;71:711–6.
- 164. Wu ML, Whittemore AS, Paffenbarger Jr RS, et al. Personal and environmental characteristics related to epithelial ovarian cancer. I. Reproductive and menstrual events

- and oral contraceptive use. Am J Epidemiol 1988;128: 1216–27.
- 165. Joly DJ, Lilienfeld AM, Diamond EL, Bross ID. An epidemiologic study of the relationship of reproductive experience to cancer of the ovary. Am J Epidemiol 1974:99:190–209.
- 166. Yen SS. Endocrinology of pregnancy. In: Creasy RK, Resnik R, editors. Maternal-fetal medicine: principles and practice. Philadelphia: Saunders; 1994. p. 382–412.
- King RJ. Biology of female sex hormone action in relation to contraceptive agents and neoplasia. Contraception 1991:43:527–42.
- 168. Rosenberg L, Palmer JR, Zauber AG, et al. A case-control study of oral contraceptive use and invasive epithelial ovarian cancer. *Am J Epidemiol* 1994;139:654–61.
- Lacey Jr JV, Mink PJ, Lubin JH, et al. Menopausal hormone replacement therapy and risk of ovarian cancer. J Am Med Assoc 2002;288:334–41.
- 170. Anderson GL, Judd HL, Kaunitz AM, et al. Effects of estrogen plus progestin on gynecologic cancers and associated diagnostic procedures: the women's health initiative randomized trial. *J Am Med Assoc* 2003;**290**:1739–48.
- 171. Riman T, Dickman PW, Nilsson S, et al. Hormone replacement therapy and the risk of invasive epithelial ovarian cancer in Swedish women. *J Natl Cancer Inst* 2002;94:497–504.
- 172. McNatty KP, Makris A, Reinhold VN, De Grazia C, Osathanondh R, Ryan KJ. Metabolism of androstenedione by human ovarian tissues in vitro with particular reference to reductase and aromatase activity. Steroids 1979;34:429–43.
- 173. Judd HL, Judd GE, Lucas WE, Yen SS. Endocrine function of the postmenopausal ovary: concentration of androgens and estrogens in ovarian and peripheral vein blood. *J Clin Endocrinol Metab* 1974;39:1020–4.
- 174. Lau KM, Mok SC, Ho SM. Expression of human estrogen receptor-alpha and -beta, progesterone receptor, and androgen receptor mRNA in normal and malignant ovarian epithelial cells. Proc Natl Acad Sci USA 1999;96:5722–7.
- 175. Godwin AK, Testa JR, Handel LM, et al. Spontaneous transformation of rat ovarian surface epithelial cells: association with cytogenetic changes and implications of repeated ovulation in the etiology of ovarian cancer. *J Natl Cancer Inst* 1992;84:592–601.
- 176. Browning MC, Anderson J. Effect of oral contraceptives on plasma testosterone concentration. Br Med J 1977;1:107.
- 177. Murphy A, Cropp CS, Smith BS, Burkman RT, Zacur HA. Effect of low-dose oral contraceptive on gonadotropins, androgens, and sex hormone binding globulin in nonhirsute women. Fertil Steril 1990;53:35–9.
- 178. Van der Vange N, Blankenstein MA, Kloosterboer HJ, Haspels JH, Thijssen JH. Effects of seven low-dose combined oral contraceptives on sex hormone binding globulin, corticosteroid binding globulin, total and free testosterone. Contraception 1990;41:345–52.
- 179. Kuhnz W, Sostarek D, Gansau C, Louton T, Mahler M. Single and multiple administration of a new triphasic oral contraceptive to women: pharmacokinetics of ethinyl estradiol and free and total testosterone levels in serum. Am J Obstet Gynecol 1991;165:596–602.
- 180. Helzlsouer KJ, Alberg AJ, Gordon GB, et al. Serum gonadotropins and steroid hormones and the development of ovarian cancer. J Am Med Assoc 1995;274:1926–30.
- Insler V, Lunenfeld B. Pathophysiology of polycystic ovarian disease: new insights. Hum Reprod 1991;6:1025–9.
- Abdel Gadir A, Khatim MS, Mowafi RS, Alnaser HM, Muharib RW, Shaw RW. Implications of ultrasonically diagnosed

- polycystic ovaries. I. Correlations with basal hormonal profiles. *Hum Reprod* 1992;7:453–7.
- 183. Robinson S, Rodin DA, Deacon A, Wheeler MJ, Clayton RN. Which hormone tests for the diagnosis of polycystic ovary syndrome? Br J Obstet Gynaecol 1992;99:232–8.
- 184. Schildkraut JM, Schwingl PJ, Bastos E, Evanoff A, Hughes C. Epithelial ovarian cancer risk among women with polycystic ovary syndrome. Obstet Gynecol 1996;88:549–54.
- 185. Cottreau CM, Ness RB, Modugno F, Allen GO, Goodman MT. Endometriosis and its treatment with danazol or lupron in relation to ovarian cancer. Clin Cancer Res 2003:9:5142–4
- 186. Harada T, Iwabe T, Terakawa N. Role of cytokines in endometriosis. Fertil Steril 2001;76:1–10.
- Nothnick WB. Treating endometriosis as an autoimmune disease. Fertil Steril 2001;76:223–31.
- 188. Gazvani R, Templeton A. New considerations for the pathogenesis of endometriosis. *Int J Gynaecol Obstet* 2002;**76**:117–26.
- 189. Oosterlynck DJ, Cornillie FJ, Waer M, Vandeputte M, Koninckx PR. Women with endometriosis show a defect in natural killer activity resulting in a decreased cytotoxicity to autologous endometrium. Fertil Steril 1991;56:45–51.
- 190. Wilson TJ, Hertzog PJ, Angus D, Munnery L, Wood EC, Kola I. Decreased natural killer cell activity in endometriosis patients: relationship to disease pathogenesis. Fertil Steril 1994;62:1086–8.
- 191. Ho HN, Chao KH, Chen HF, Wu MY, Yang YS, Lee TY. Peritoneal natural killer cytotoxicity and CD25+ CD3+ lymphocyte subpopulation are decreased in women with stage III–IV endometriosis. Hum Reprod 1995;10:2671–5.
- 192. Kikuchi Y, Ishikawa N, Hirata J, Imaizumi E, Sasa H, Nagata I. Changes of peripheral blood lymphocyte subsets before and after operation of patients with endometriosis. Acta Obstet Gynecol Scand 1993;72:157–61.
- 193. Haney AF, Muscato JJ, Weinberg JB. Peritoneal fluid cell populations in infertility patients. Fertil Steril 1981;35:696–8.
- 194. Halme J, Becker S, Hammond MG, Raj S. Pelvic macrophages in normal and infertile women: the role of patent tubes. Am J Obstet Gynecol 1982;142:890–5.
- Halme J, Becker S, Wing R. Accentuated cyclic activation of peritoneal macrophages in patients with endometriosis. Am J Obstet Gynecol 1984;148:85–90.
- Halme J, Becker S, Haskill S. Altered maturation and function of peritoneal macrophages: possible role in pathogenesis of endometriosis. Am J Obstet Gynecol 1987;156:783–9.
- 197. Dunselman GA, Hendrix MG, Bouckaert PX, Evers JL. Functional aspects of peritoneal macrophages in endometriosis of women. *J Reprod Fertil* 1988;82:707–10.
- 198. Braun DP, Muriana A, Gebel H, Rotman C, Rana N, Dmowski WP. Monocyte-mediated enhancement of endometrial cell proliferation in women with endometriosis. Fertil Steril 1994:61:78–84.
- 199. Oosterlynck DJ, Meuleman C, Waer M, Koninckx PR. Transforming growth factor-beta activity is increased in peritoneal fluid from women with endometriosis. Obstet Gynecol 1994;83:287–92.
- 200. Akoum A, Lemay A, Paradis I, Rheault N, Maheux R. Secretion of interleukin-6 by human endometriotic cells and regulation by proinflammatory cytokines and sex steroids. Hum Reprod 1996;11:2269–75.
- 201. Tseng JF, Ryan IP, Milam TD, et al. Interleukin-6 secretion in vitro is upregulated in ectopic and eutopic endometrial stromal cells from women with endometriosis. *J Clin Endocrinol Metab* 1996;**81**:1118–22.
- 202. Tsudo T, Harada T, Iwabe T, et al. Altered gene expression and secretion of interleukin-6 in stromal cells derived from endometriotic tissues. Fertil Steril 2000;73:205–11.

- 203. Kharfi A, Boucher A, Akoum A. Abnormal interleukin-1 receptor type II gene expression in the endometrium of women with endometriosis. Biol Reprod 2002;66:401–6.
- 204. McLaren J, Prentice A, Charnock-Jones DS, Smith SK. Vascular endothelial growth factor (VEGF) concentrations are elevated in peritoneal fluid of women with endometriosis. Hum Reprod 1996;11:220–3.
- Oosterlynck DJ, Meuleman C, Sobis H, Vandeputte M, Koninckx PR. Angiogenic activity of peritoneal fluid from women with endometriosis. Fertil Steril 1993;59:778–82.
- 206. Rodgers WH, Osteen KG, Matrisian LM, Navre M, Giudice LC, Gorstein F. Expression and localization of matrilysin, a matrix metalloproteinase, in human endometrium during the reproductive cycle. Am J Obstet Gynecol 1993;168:253–60.
- 207. Gleicher N, el-Roeiy A, Confino E, Friberg J. Is endometriosis an autoimmune disease? *Obstet Gynecol* 1987;**70**:115–22.
- Sinaii N. Autoimmune and related diseases among women with endometriosis: a survey analysis. Hum Reprod 2002;17:2715–24.
- Startseva NV. Clinical immunological aspects of genital endometriosis. Akush Ginekol (Mosk) 1980;3:23–6.
- Weed JC, Arquembourg PC. Endometriosis: can it produce an autoimmune response resulting in infertility? Clin Obstet Gynecol 1980;23:885–93.
- 211. Mathur S, Peress MR, Williamson HO, et al. Autoimmunity to endometrium and ovary in endometriosis. Clin Exp Immunol 1982;50:259–66.
- 212. el-Roeiy A, Dmowski WP, Gleicher N, et al. Danazol but not gonadotropin-releasing hormone agonists suppresses autoantibodies in endometriosis. Fertil Steril 1988;50:864–71.
- 213. Kennedy SH, Starkey PM, Sargent IL, Hicks BR, Barlow DH. Antiendometrial antibodies in endometriosis measured by an enzyme-linked immunosorbent assay before and after treatment with danazol and nafarelin. Obstet Gynecol 1990;75:914–8.
- 214. Imai A, Takagi A, Tamaya T. Gonadotropin-releasing hormone analog repairs reduced endometrial cell apoptosis in endometriosis in vitro. Am J Obstet Gynecol 2000;182:1142–6.
- 215. Cramer DW, Missmer SA. The epidemiology of endometriosis. *Ann NY Acad Sci* 2002;**955**:11–22.
- Cramer DW, Wilson E, Stillman RJ, et al. The relation of endometriosis to menstrual characteristics, smoking, and exercise. J Am Med Assoc 1986;255:1904

 –8.
- 217. Matorras R, Rodiquez F, Pijoan JI, Ramon O, Gutierrez de Teran G, Rodriguez-Escudero F. Epidemiology of endometriosis in infertile women. Fertil Steril 1995;63:34–8.
- 218. Moen MH, Schei B. Epidemiology of endometriosis in a Norwegian county. Acta Obstet Gynecol Scand 1997;76:559–62.
- 219. Berube S, Marcoux S, Maheux R. Characteristics related to the prevalence of minimal or mild endometriosis in infertile women. Canadian Collaborative Group on Endometriosis. *Epidemiology* 1998;9:504–10.
- Grodstein F, Goldman MB, Ryan L, Cramer DW. Relation of female infertility to consumption of caffeinated beverages. Am J Epidemiol 1993;137:1353–60.
- 221. Parazzini F, La Vecchia C, Negri E, Gentile A. Menstrual factors and the risk of epithelial ovarian cancer. *J Clin Epidemiol* 1989;42:443–8.
- 222. Sangi-Haghpeykar H. Epidemiology of endometriosis to menstrual characteristics, smoking, and exercise. *J Am Med Assoc* 1986;255:1904–8.
- 223. Obermeyer CM, Armenian HK, Azoury R. Endometriosis in Lebanon. Am J Epidemiol 1986;124:762–5.
- 224. Parazzini F, Ferraroni M, Fedele L, Bocciolone L, Rubessa S, Riccardi A. Pelvic endometriosis: reproductive and menstrual risk factors at different stages in Lombardy, northern Italy. *J Epidemiol Community Health* 1995;49:61–4.

- 225. Gruppo Italiano per lo Studio dell' endometriosi. Risk factors for pelvic endometriosis in women with pelvic pain or infertility. Eur J Obstet Gynecol Reprod Biol 1999;83:195–99.
- 226. Signorello LB, Harlow BL, Cramer DW, Spiegelman D, Hill JA. Epidemiologic determinants of endometriosis: a hospital-based case-control study. Ann Epidemiol 1997;7:267–74.
- 227. Hadfield RM, Yudkin PL, Coe CL, et al. Risk factors for endometriosis in the rhesus monkey (Macaca mulatta): a case-control study. Hum Reprod Update 1997;3:109–15.
- 228. Zeitoun KM, Bulun SE. Aromatase: a key molecule in the pathophysiology of endometriosis and a therapeutic target. Fertil Steril 1999;72:961–9.
- 229. Bulun SE, Mahendroo MS, Simpson ER. Polymerase chain reaction amplification fails to detect aromatase cytochrome P450 transcripts in normal human endometrium or decidua. *J Clin Endocrinol Metab* 1993;**76**:1458–63.
- 230. Casey ML, MacDonald PC. Origin of estrogen and the regulation of estrogen formation in postmenopausal women. In: Buchsbaum JH, editor. *The menopause*. New York: Springer-Verlag; 1983. p. 1–8.
- 231. MacDonald PC, Rombaut RP, Siiteri PK. Plasma precursors of estrogen. I. Extent of conversion of plasma delta-4-androstenedione to estrone in normal males and nonpregnant normal, castrate and adrenalectomized females. J Clin Endocrinol Metab 1967;27:1103–11.
- 232. Wu L, Einstein M, Geissler WM, Chan HK, Elliston KO, Andersson S. Expression cloning and characterization of human. *J Biol Chem* 1993;268:12964–9.
- 233. Labrie Y, Durocher F, Lachance Y, et al. The human type II 17 beta-hydroxysteroid dehydrogenase gene encodes two alternatively spliced mRNA species. DNA Cell Biol 1995;14:849–61.
- 234. Andersson S, Moghbrabi N. Physiology and molecular genetics of 17 beta-hydroxysteroid dehydrogenases. *Steroids* 1997;**62**:143–7.
- 235. Zeitoun K, Takayama K, Sasano H, et al. Deficient 17beta-hydroxysteroid dehydrogenase type 2 expression in endometriosis: failure to metabolize 17 beta-estradiol. J Clin Endocrinol Metab 1998;83:4474–80.
- 236. Halme J, Stovall D. Endometriosis and its medical management. In: Wallach EE, Zacur HA, editors. Reproductive medicine and surgery. St Louis: Mosby; 1995. p. 692–710.
- Osteen KG, Bruner KL, Sharpe-Timms KL. Steroid and growth factor regulation of matrix metalloproteinase expression and endometriosis. Semin Reprod Endocrinol 1996;14:247–55.
- 238. Bruner-Tran KL, Rier SE, Eisenberg E, Osteen KG. The potential role of environmental toxins in the pathophysiology of endometriosis. *Gynecol Obstet Invest* 1999;48:45–56.
- 239. Huang JC, Liu DY, Yadollahi S, Wu KK, Dawood MY. Interleukin-1 beta induces cyclooxygenase-2 gene expression in cultured endometrial stromal cells. *J Clin Endocrinol Metab* 1998;83:538–41.
- 240. Noble LS, Takayama K, Zeitoun KM, et al. Prostaglandin E2 stimulates aromatase expression in endometriosis-derived stromal cells. J Clin Endocrinol Metab 1997;82:600–6.
- 241. Kitawaki J, Noguchi T, Amatsu T, et al. Expression of aromatase cytochrome P450 protein and messenger ribonucleic acid in human endometriotic and adenomyotic tissues but not in normal endometrium. Biol Reprod 1997;57:514–9.
- 242. Reed MJ. The discriminant-function test: a marker of Th1/ Th2 cell cytokine secretion and breast tumor estrogen synthesis. Mol Med Today 1995;1:98–103.
- 243. Reed MJ, Purohit A. Breast cancer and the role of cytokines in regulating estrogen synthesis: an emerging hypothesis. *Endocr Rev* 1997;18:701–15.

- 244. Macdiarmid F, Wang D, Duncan LJ, Purohit A, Ghilchick MW, Reed MJ. Stimulation of aromatase activity in breast fibroblasts by tumor necrosis factor alpha. Mol Cell Endocrinol 1994;106:17–21.
- 245. Adams EF, Rafferty B, White MC. Interleukin 6 is secreted by breast fibroblasts and stimulates 17 beta-oestradiol oxidoreductase activity of MCF-7 cells: possible paracrine regulation of breast 17 beta-oestradiol levels. Int J Cancer 1991;49:118–21.
- 246. Duncan LJ, Coldham NG, Reed MJ. The interaction of cytokines in regulating oestradiol 17 beta-hydroxysteroid dehydrogenase activity in MCF-7 cells. *J Steroid Biochem Mol Biol* 1994;49:63–8.
- 247. Purohit A, Duncan LJ, Wang DY, Coldham NG, Ghilchick MW, Reed MJ. Paracrine control of oestrogen production in breast cancer. Endocr Relat Cancer 1997;4:323–30.
- 248. Singer CF, Kronsteiner N, Hudelist G. Interleukin 1 system and sex steroid receptor expression in human breast cancer: interleukin 1alpha protein secretion is correlated with malignant phenotype. Clin Cancer Res 2003;9:4877–83.
- 249. Danforth Jr DN, Sgagias MK. Interleukin 1 alpha blocks estradiol-stimulated growth and downregulates the estrogen receptor in MCF-7 breast cancer cells in vitro. *Cancer Res* 1991;51:1488–93.
- 250. Runesson E, Ivarsson K, Janson PO, Brannstrom M. Gonadotropin- and cytokine-regulated expression of the chemokine interleukin 8 in the human preovulatory follicle of the menstrual cycle. J Clin Endocrinol Metab 2000;85:4387–95.
- 251. Tabibzadeh S. Cytokines and the hypothalamic-pituitaryovarian-endometrial axis. Hum Reprod 1994;9:947–67.
- 252. Ray A, Prefontaine KE, Ray P. Down-modulation of interleukin-6 gene expression by 17 beta-estradiol in the absence of high affinity DNA binding by the estrogen receptor. J Biol Chem 1994;269:12940–6.
- 253. Tazuke SI, Giudice LC. Growth factors and cytokines in endometrium, embryonic development, and maternal: embryonic interactions. Semin Reprod Endocrinol 1996;14:231–45.
- 254. Berg G, Ekerfelt C, Hammar M, Lindgren R, Matthiesen L, Ernerudh J. Cytokine changes in postmenopausal women treated with estrogens: a placebo-controlled study. Am J Reprod Immunol 2002;48:63–9.
- 255. Saucedo R, Rico G, Basurto L, Ochoa R, Zarate A. Transdermal estradiol in menopausal women depresses interleukin-6 without affecting other markers of immune response.

 Gynecol Obstet Invest 2002;53:114–7.
- 256. Walsh BW, Cox DA, Sashegyi A, Dean RA, Tracy RP, Anderson PW. Role of tumor necrosis factor-alpha and interleukin-6 in the effects of hormone replacement therapy and raloxifene on C-reactive protein in postmenopausal women. Am J Cardiol 2001;88:825–8.
- 257. Straub RH, Hense HW, Andus T, Scholmerich J, Riegger GA, Schunkert H. Hormone replacement therapy and interrelation between serum interleukin-6 and body mass index in postmenopausal women: a population-based study. J Clin Endocrinol Metab 2000;85:1340–4.
- 258. Purohit A, Reed MJ. Regulation of estrogen synthesis in postmenopausal women. *Steroids* 2002;**67**:979–83.
- 259. Purohit A, Singh A, Ghilchik MW, Reed MJ. Inhibition of tumor necrosis factor alpha-stimulated aromatase activity by microtubule-stabilizing agents, paclitaxel and 2-methoxyestradiol. Biochem Biophys Res Commun 1999;261:214–7.
- Hillier SG, Tetsuka M. An anti-inflammatory role for glucocorticoids in the ovaries? J Reprod Immunol 1998;39:21–7.
- Ziltener HJ, Maines-Bandiera S, Schrader JW, Auersperg N. Secretion of bioactive interleukin-1, interleukin-6, and

- colony-stimulating factors by human ovarian surface epithelium. *Biol Reprod* 1993;**49**:635–41.
- Adashi EY. The potential role of interleukin-1 in the ovulatory process: an evolving hypothesis. Mol Cell Endocrinol 1998;140:77–81.
- 263. Smith G, Roberts R, Hall C, Nuki G. Reversible ovulatory failure associated with the development of luteinized unruptured follicles in women with inflammatory arthritis taking non-steroidal anti-inflammatory drugs. Br J Rheumatol 1996;35:458–62.
- 264. Subbaramaiah K, Zakim D, Weksler BB, Dannenberg AJ. Inhibition of cyclooxygenase: a novel approach to cancer prevention. Proc Soc Exp Biol Med 1997;216: 201–210.
- 265. Mandlekar S, Kong AN. Mechanisms of tamoxifen-induced apoptosis. Apoptosis 2001;6:469–77.

- 266. Rodriguez GC, Nagarsheth NP, Lee KL, et al. Progestin-induced apoptosis in the Macaque ovarian epithelium: differential regulation of transforming growth factor-beta. J Natl Cancer Inst 2002;94:50–60.
- 267. Adashi EY. The potential relevance of cytokines to ovarian physiology: the emerging role of resident ovarian cells of the white blood cell series. *Endocr Rev* 1990;11:454–64.
- 268. Fukuoka M, Yasuda K, Fujiwara H, Kanzaki H, Mori T. Interactions between interferon gamma, tumour necrosis factor alpha, and interleukin-1 in modulating progesterone and oestradiol production by human luteinized granulosa cells in culture. Hum Reprod 1992;7:1361–4.
- 269. Yan Z, Hunter V, Weed J, Hutchison S, Lyles R, Terranova P. Tumor necrosis factor-alpha alters steroidogenesis and stimulates proliferation of human ovarian granulosal cells in vitro. Fertil Steril 1993;59:332–8.